

2016

# Addressing the mismatch between restoration objectives and monitoring needs to support mangrove management

Debra J. Stokes  
*Southern Cross University*

Richard H. Bulmer  
*University of Auckland*

Carolyn J. Lundquist  
*University of Auckland*

---

## Publication details

Postprint of: Stokes, DJ, Bulmer, RH & Lundquist, CJ 2016, 'Addressing the mismatch between restoration objectives and monitoring needs to support mangrove management', *Ocean & Coastal Management*, vol. 134, pp. 69-78.

Published version available from:

<http://dx.doi.org/10.1016/j.ocecoaman.2016.09.024>

ePublications@SCU is an electronic repository administered by Southern Cross University Library. Its goal is to capture and preserve the intellectual output of Southern Cross University authors and researchers, and to increase visibility and impact through open access to researchers around the world. For further information please contact [epubs@scu.edu.au](mailto:epubs@scu.edu.au).

## 1. Introduction

Estuaries bridge our terrestrial and marine environments; they are unique and diverse ecosystems that provide invaluable ecosystem services (Barbier et al., 2011). Saltmarsh, mangrove and seagrass vegetation are critical vegetated habitats in estuaries, providing food and protection for numerous estuarine species, including fish, bivalves, crustaceans and birds (Thrush et al., 2013). Unfortunately, estuaries are also sites where impacts from human activity are often amplified, deeming them some of the most heavily used and threatened systems on the planet (Lotze et al., 2006, Halpern et al., 2008) planet. For example, the huge growth in aquaculture has resulted in an enormous loss of coastal/wetland vegetation, with shrimp farming contributing to some ~ 38 % of global mangrove loss and other aquaculture has contributed to a further 14 % loss (Ellison, 2008). Other direct impacts may be driven by land reclamation, dredging, port development and coastal urbanisation, while pollutants such as nutrients, metals and plastics from the surrounding catchment can accumulate within estuaries and harbours (Kennish, 2002). Such impacts can trigger dieback or profound shifts in the composition and health of coastal vegetation and other estuarine habitats (Thrush et al., 2003, Saintilan et al., 2014, Doughty et al., 2015). Degradation of coastal habitats can be addressed by removing threatening processes, however often restoration and rehabilitation is required to support ecosystem recovery (Ferrier and Jenkins, 2010).

Wetland restoration is a common management strategy applied around the globe, although most of this work is taking place in developed countries (see Bayraktarov et al., 2016 for review). A key objective is to restore ecosystem services and halt further loss of vegetation (Zhao et al., 2016). A recent review of 235 coastal restoration projects analysed the successes and costs, all of which involved some form of replanting or reseedling for restoration of coral reefs, seagrass, mangroves, saltmarshes or oyster reefs (Bayraktarov et al., 2016). In temperate regions of north America, replanting of saltmarsh has been central to the success of various restoration projects (Zedler, et al., 2012). Elsewhere, mangrove rehabilitation programs have been implemented to restore forest cover and habitat functionality (Milbrandt et al., 2015, Osland et al., 2012). Rarely has removal of native vegetation been considered a restoration technique, however in recent times resource management agencies in New Zealand have removed mangroves in an attempt to restore tidal flat ecosystems (Harty, 2009; Morrisey et al., 2010; Lundquist et al., 2014).

Despite a substantial global decline in mangrove distribution (Giri et al., 2011), many temperate mangrove forests are increasing in distribution (Morrisey et al., 2010). In New Zealand, this typically occurs where sediment loads are high and mangroves colonise seaward across bare mudflats (Lundquist et al., 2014, Swales et al., 2015, Stokes et al., 2010). In southern Australia drought and limited sediment loads (leading to compaction) are suggested causes of landward colonisation by mangroves (Rogers et al., 2005; Saintilan and Rogers, 2013).

When discussing the various approaches to managing mangroves out of coastal and estuarine habitats, Elliot et al. (2007) suggest that for recovery to be truly successful, the community established has to be similar in species composition, population density and size and biomass structure to that which was present prior to mangroves, or similar to that described at a site where mangroves are not present. Understanding how a site responds to the eradication of some or all of its mangroves is vital if we are to develop and implement effective habitat management strategies. There is limited reporting on the impacts of mangrove removal activities in temperate mangrove systems, in both academic and grey literature. This limits the ability to adopt best practice mangrove management techniques that minimise adverse impacts and to identify cost-effective means to achieve restoration success. Furthermore, there is a real risk that an unsubstantiated paradigm shift will take hold in policy development whereby the removal of mangroves becomes identified as a

47 positive ecosystem service. This is a possibility if monitoring of the impacts of their removal  
48 continues to be absent or minimal, and any management actions are assumed, but may not actually  
49 achieve restoration objectives.

50 This paper reviews the monitoring programs associated with mangrove eradication in New Zealand,  
51 and identifies the key monitoring protocols that should be included in mangrove removal activities.  
52 These measures should determine whether mangrove removal leads to the desired environmental  
53 outcomes, and also to inform future coastal management. New Zealand community groups and  
54 coastal management authorities have been clearing mangroves for over a decade now (de Luca,  
55 2015). This provides us with the opportunity to explore the varied approaches to the removal of  
56 mangrove vegetation. Furthermore, we can use existing monitoring data to assess relative success  
57 of mangrove management activities to date in achieving restoration objectives.

## 58 **1.1 Background – Mangroves in New Zealand**

59 *Avicennia marina* subsp. *australasica* is the only mangrove species occurring in New Zealand  
60 (Morrisey et al., 2010). Early European records document the presence of mangroves (Swales et al.,  
61 2015) and pollen analysis confirms the presence of mangrove pollen in sedimentary deposits older  
62 than 8000 years (Mildenhall, 2001), confirming that this is an indigenous plant of the North Island  
63 of New Zealand. Due to a combination of climate gradients (Beard, 2006) and dispersal limitation  
64 (de Lange and de Lange 1994), mangroves are presently only found in harbours and bays in the  
65 northern half of the North Island, north of around latitude 38°. Phases of mangrove expansion have  
66 been mapped from aerial photographs dating back to the 1940s (see Swales et al., 2015). Rates of  
67 increase, and the periods during which colonisation was most rapid, have been variable over this  
68 time (Swales et al., 2007; Morrisey et al., 2010; Stokes et al., 2010). Drivers of mangrove expansion  
69 include estuarine infilling and associated changes in mean bed level (Ellis et al., 2004; Swales et al.,  
70 2007; Stokes et al., 2009), and decreased storm and wave activity linked to El Niño (Swales et al.,  
71 2015).

72 Reports produced in the 1970s (Chapman et al., 1976a and b) alerted authorities to dwindling  
73 mangrove habitat following land reclamation and grazing impacts. As a result, New Zealand  
74 mangroves were granted protected status under the New Zealand Coastal Policy (Harty, 2009).  
75 Local regulatory bodies such as Regional Councils operate according to the NZ Coastal Policy and  
76 Resource Management Act, and as such, any mangrove clearing must be approved via the resource  
77 consent process (Harty, 2009). However, minimal monitoring information is available from prior  
78 mangrove removal activities to inform decision making with respect mangrove removals. In the New  
79 Zealand context, natural resource management falls under the jurisdiction of district and regional  
80 councils (Harty, 2009).

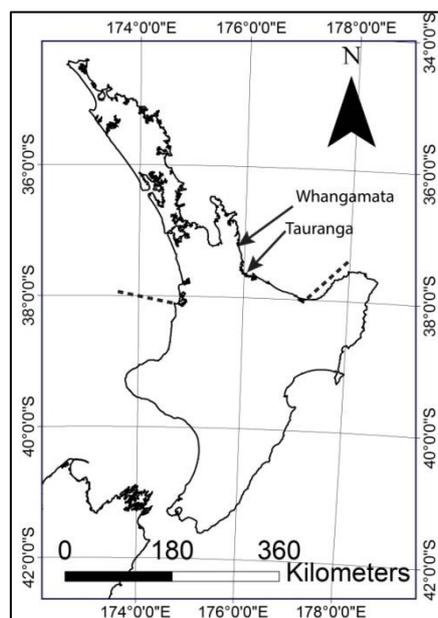
81 Interestingly, a recent re-evaluation of the NZ Coastal Policy (2010) has altered the status of  
82 mangroves, in which the policy statement “no longer identifies mangroves as a species worthy of  
83 conservation”. In line with this change, some regional councils have updated their regional policies.  
84 For example, Northland Regional Council have described conditions under which mangrove removal  
85 can be undertaken as a ‘discretionary activity’ (i.e. do not require the full consent process for  
86 approval, and does not require monitoring of impacts). Waikato Regional Council continues to  
87 require consent application for mangrove removal, and they are involved in large consented  
88 clearings in both Whangamata and Tairua Harbours. Detailed monitoring has been undertaken in  
89 Whangamata Harbour (see Bulmer and Lundquist, 2015 and 2016 for monitoring results). Auckland  
90 Council and Bay of Plenty Regional Council have draft plans that are relatively permissive for removal  
91 of mangroves and seedlings (DeLuca, 2015), although the extent of the removal is bounded by an

92 arbitrary historical mangrove boundary (i.e. back to where mangroves fringed the site in 1996).  
93 Recently (August 2016), the proposed 1996 mangrove boundary for the Auckland coastal plan was  
94 removed after consultation hearings, and the final plan is expected to return to less permissive  
95 mangrove removal conditions.

96 Mangrove clearing across New Zealand has been undertaken both with consent (and so legally), and  
97 without consent (illegally). A number of methods have been applied to clear the above-ground  
98 mangrove vegetation, with earlier work mostly done using chainsaws and/or slash cutters,  
99 supported by the hand-pulling of seedlings across the seaward mud/sandflats. The size of any area  
100 cleared at one time was mostly limited by the manpower, and of those activities that were  
101 documented, the extent of removals ranged from  $< 0.5$  ha to  $\sim 2$  ha. More recently, the  
102 introduction of wide-track machinery has resulted in larger areas being cleared at any one time. The  
103 largest was cleared in Tauranga Harbour in 2010, which totalled approximately 101 ha, however  
104 cleared plots within the sub-estuaries ranged in size from 1.5 ha to 20 ha (see Lundquist et al., 2014  
105 for a more comprehensive list of clearing activities).

106 The treatment of the mangrove debris has also varied across management jurisdictions. Hand-  
107 cleared vegetation has often been stockpiled on site, and incinerated once dried. Some hand-  
108 cleared material was removed from the intertidal zone which required access for a truck or tractor  
109 to transport the material off-site. In 2010, vegetation which had been cleared with wide-track  
110 machinery, was subsequently mulched and deposited on the intertidal mudflat, resulting in a  
111 persistent blanket of debris (Lundquist et al., 2014).

112 We summarise national monitoring of mangrove removals, focussing on three detailed case studies  
113 of legally consented mangrove removal in New Zealand (see Figure 1). Firstly we identify what  
114 specific information is needed to inform sustainable management of mangrove habitats, and  
115 secondly to determine whether the desired management objectives for mangrove removal were  
116 detected by monitoring. We then present monitoring variables that have been successfully used to  
117 elucidate success at achieving restoration objectives in mangrove management.



118

119 Figure 1. Location of Whangamata Harbour and Tauranga Harbour, on the north island of New  
120 Zealand. The dashed line marks the approximate southern extent of mangrove habitation.

121 **2. Case Studies**

122 Management objectives for mangrove removal in New Zealand generally cite two key desired  
123 outcomes: a) to return a site to sandy benthic habitat; and b) to see a return of benthic communities  
124 that were present prior to mangrove presence, with healthy bivalve abundance (Stokes 2010). As  
125 such, monitoring designs have tended to focus on these attributes. Limited long term studies are  
126 available to support any attempt at predicting the scale of the potential impacts of mangrove  
127 removal and the rate of recovery. To date, detailed studies of physical and ecological changes are  
128 mostly limited to 2 or 3 years post-clearance. What can be concluded from these studies, however,  
129 is that each site responds differently according to exposure to tidal currents, agitation of sediments  
130 from wind and waves, the depth of muddy material, the size of the cleared area and the removal  
131 methods used (Park, 2004; Alfaro, 2010; Stokes et al., 2010; Lundquist et al., 2014)

132 **2.1 Case study 1(a) – Tauranga Harbour: mangrove clearance by community groups**

133 Tauranga Harbour is a large barrier estuary covering 200 km<sup>2</sup>. Numerous small sub-estuaries nestle  
134 into steep sub-catchments that introduce considerable sediment loads to the harbour. The city of  
135 Tauranga wraps around the margins of the southern section with a population of around 100,000.  
136 By 2003, mangrove coverage in some of these sub-estuaries increased from narrow marginal fringes  
137 in the 1950s (Park 2004) to occupy up to 38 % of each embayment, totalling more than 500 hectares  
138 across the Harbour.

139 After some unconsented clearing of mangroves during the 1980s and 1990s, the regional council  
140 (Bay of Plenty Regional Council– BoPRC) and the local District/City council supported the creation of  
141 ‘Estuary Care Groups’ to facilitate a more regulated approach to mangrove management. Using  
142 volunteer labour these groups removed < 20 hectares of mangroves from the harbour in the space  
143 of 5 - 10 years. The Regional Council also provided groups with community monitoring guidelines to  
144 evaluate surface elevation changes, surface macrofauna communities and intertidal vegetation  
145 boundaries (Schwarzet al., 2004). The resulting data were delivered to the regulatory bodies  
146 (council), not only as evidence of on-going mangrove expansion but also to support consent  
147 applications for further clearance of mangroves. Unfortunately the results and analysis undertaken  
148 by these care groups are not readily available or useful for two reasons, firstly that substantial  
149 inconsistencies in methodologies occurred in this volunteer monitoring, both in methods and in  
150 regularity of monitoring, and secondly that poor documentation and record keeping at council level  
151 occurred such that results are not available to inform further management activities.

152 The extent of mangrove removal undertaken by community groups at this time was limited by the  
153 amount of volunteer labour available to each group, and the limitations of the methods employed  
154 (mainly chain saws and brushcutters which were used to partially clear mangrove stands). Consent  
155 conditions stipulated that only mangroves which had established since 1984 could be cleared, while  
156 older stands were to remain (Stokes, 2010).

157 While detailed results of this activity have not yet been compiled by council or any data made  
158 available, independent postgraduate research monitored changed over an 18 month period within  
159 one of the sub-estuaries. The study reported a coarsening of surface sediment, in the first 12  
160 months after mangrove removal coupled with a marked fall in surface topography of up to 38 mm.  
161 In an analysis of benthic macroinvertebrates no links between community structure and ‘restoration’  
162 could be confirmed, at least in the 1 mm sieve size fraction (Stokes, 2010).

163 **2.2 Case Study 1(b): machinery assisted clearing in Tauranga Harbour**

164 In 2010, Bay of Plenty Regional Council (BoPRC) authorised the use of wide-track machinery to clear  
165 approximately 110 hectares of mangroves from 10 sub-estuaries of Tauranga Harbour. Vegetation  
166 was mulched as it was cleared, and the mulchate was left in-situ (Lundquist et al., 2014). Under the  
167 issued consent, no collection of baseline data was required prior to this large-scale clearance  
168 (Consent 66505 and 65693). The consent was based on a small trial of mechanical clearing, whereby  
169 epifauna were observed to rapidly recolonise tracks within a tidal cycle (Park 2008, in Harrison  
170 Grierson Consultants Ltd, Resource Consent Application 2012); however, the tracking and mulching  
171 methods that did occur during consent works had little correspondence with methods tested in the  
172 small trial. Post-clearing monitoring required by the resource consent was limited to qualitative  
173 observations, with site visits required 'every two days for no less than 1 week and up to two  
174 months'. No formal technical reports or data was collected, as these visits were primarily interested  
175 in the lack of dispersal of mulchate, and thus, lack of likely impact on neighbouring habitats.  
176 Photographs were taken to record evidence of wading bird activity, distribution of mulch material,  
177 and changes to sediment characteristics. Later, after concerns were raised by community groups,  
178 further monitoring was initiated annually in two of the ten estuaries exposed to mulching.

179 Due to concerns by independent scientists at the National Institute of Water and Atmospheric  
180 Research about the scale of the mangrove removals in the consent, and the lack of data to inform  
181 the consent activity, independent research was initiated. This included the collection of baseline  
182 samples prior to mangrove removals, to gather detailed data on the impacts of persistent mulchate.  
183 The study assessed the general condition of three of the ten sub-estuaries, though delays in consent  
184 activities meant that sampling only occurred within two sub-estuaries. Transect surveys of  
185 macrofauna and sediment characteristics were undertaken at 3, 6 and 12 months post-clearing in  
186 both estuaries. Additional sampling by NIWA, as part of a separate research project, occurred in  
187 five sub-estuaries approximately three years after removals. In both studies, visual surveys of sites  
188 at positions within mulch zones, and in adjacent mangrove and adjacent tidal flats were combined  
189 with sediment cores (grainsize, organic content, chlorophyll a) and macrofaunal cores to interpret  
190 both recovery trajectories and document adverse impacts. Early results identified limited dispersal  
191 or decomposition of the mulch material which resulted in anoxic sediments, elevated levels of  
192 phosphorous and ammonium, algal blooms and some occurrences of lowered dissolved oxygen  
193 below ANZECC guidelines (Lundquist et al., 2012). No evidence of site recovery was found within 12  
194 months at either site, and sites all exhibited degraded conditions at 3 years post removal (Lundquist,  
195 unpublished data).

196

### 197 **2.3 Case Study 2: Whangamata Harbour**

198 In 2012, Waikato Regional Council (WRC), taking note of the Tauranga Harbour experience,  
199 approved clearance of up to 23 hectares of mangroves in Whangamata Harbour. Consent conditions  
200 required detailed baseline surveys and regular monitoring for up to three years post-clearance  
201 (Bulmer and Lundquist 2014, 2015, 2016). WRC chose to trial and monitor both mechanical and  
202 hand removal methods, at locations with varying degrees of tidal flushing and exposure to wind  
203 waves and storm conditions. Here, however, all mulchate was stockpiled and incinerated rather  
204 than spread across the cleared substrate (Bulmer and Lundquist, 2014).

205 To date, these monitoring conditions are the most detailed required by any mangrove removal  
206 consent in New Zealand. As such, it is likely that a better estimate of the timeframes required to  
207 achieve a state of restoration will become possible. In addition, rather than simply reporting on  
208 point-in-time physical and biological conditions, the monitoring program includes an assessment of

209 trends and triggers. This approach provides an opportunity to adopt an adaptive management  
 210 approach whereby further clearances can be discontinued if any harmful trends are identified, or  
 211 site-specific remediation can be undertaken should it be deemed appropriate. Importantly, this  
 212 approach attempts to quantify recovery or restoration using a range of physical and biological  
 213 parameters. Criteria included in the monitoring at Whangamata Harbour are summarised in Table 1.

214 Table 1. Key criteria used to identify 3 year trends, characterised as positive or negative trends.

CRITERIA	POSITIVE TREND & TRIGGERS	NEGATIVE TREND & TRIGGERS
Sediment characteristics (mud content)	decrease in mud content; increase in Redox Potential Depth (RDP)	increase in mud content of > 5 % and/or RPD decreases by > 10 %
Sediment characteristics (compaction)	compaction class (firm, slightly soft, soft, very soft) trending towards more compact sediments	A decrease in compaction relative to initial conditions
Saltmarsh and bank erosion	No significant change to bank condition	Significant erosion (characterised on a case-by-case basis)
Mangrove encroachment	No significant change in saltmarsh-mangrove boundary	Landward transgression in the saltmarsh-mangrove boundary of more than 30 cm in a 12 month period
Seagrass extent and density	Increase or no significant reduction in seagrass cover (defined as > 10 % areal coverage in any 3 month period and > 20 % in any 12 month period)	Decreases in areal coverage > 20 % over twelve months (noting seasonal variability)
Benthic fauna	Community composition and abundance trends towards open intertidal tidal flat. > In abundance to at least 20 % of the abundance of the top 5 ranked taxa found on adjacent tidal flats	Minimal colonisation by macrofauna; presence of macroalgae blooms.

215

216 Following 24 months of monitoring, Bulmer and Lundquist (2015) reported that larger sites (greater  
 217 than 1 hectare) showed some increase in mud content, suggesting an undesirable trend (Bulmer and  
 218 Lundquist, 2015). It was unclear, however, whether this sediment deposition was related to  
 219 mangrove removals or land-based activities (such as forestry) in the neighbouring catchment.  
 220 Macrofaunal communities were more similar to intact mangrove community structure than adjacent  
 221 tidal flats and this was similar across all sites regardless of size, exposure or removal methods.  
 222 However, some trends toward sandflat communities were observed three years after removals  
 223 (Bulmer and Lundquist, 2016). The study reported a more rapid coarsening of surface sediments  
 224 where mangroves were cleared by hand, while the width of the area cleared correlated with  
 225 recovery trends. Sites where width of removal was < 30 m showed a more rapid change in sediment  
 226 characteristics, as did any cleared habitat positioned near tidal channels (and increased  
 227 hydrodynamic forces). Adverse impacts, as witnessed in Tauranga Harbour (i.e. long-lasting anoxic  
 228 sediments, macroalgal blooms and delayed colonisation by macrofauna) appeared to be minimal  
 229 across Whangamata Harbour, although trends toward sandier substrates were slower than  
 230 anticipated.

231 The case studies outlined above identify the varied approaches to monitoring associated with  
 232 mangrove removal in New Zealand. What is evident is that the numerous management authorities  
 233 have applied inconsistent approaches to mangrove removal, and most alarmingly the larger scale  
 234 removals can result in adverse impacts. The introduction of mechanical clearing techniques, and  
 235 poorly documented monitoring (post-clearance) may delay the adoption of best practice in terms of  
 236 appropriate scale of removal plots, and the timeframes identified to undertake the clearing  
 237 activities. It is important that coastal managers are aware of the potential impacts associated with  
 238 their preferred approach to mangrove management, to ensure the likelihood of negative outcomes  
 239 are mitigated (Lundquist et al., 2012, 2014). A new monitoring framework, incorporating monitoring

240 data to date, can improve the cost-effectiveness of the techniques employed. It can also provide an  
241 improved opportunity to identify and prioritise areas where restoration activities are likely to result  
242 in successful restoration after mangrove clearing.

### 243 **3. Knowledge Gaps**

244 'Ecological restoration' implies an endeavour to return an ecosystem back to, as much as possible,  
245 it's 'original' condition (Dale et al., 2014). Under this principle, the ultimate goal of wetland  
246 restoration is "to create a self-organising, self-maintaining and functioning natural ecosystem that is  
247 resilient to perturbation without further assistance" (Zhao et al., 2016). Using this definition, how  
248 do we evaluate the success of using mangrove clearing as a restoration tool? A cleared site should  
249 reflect similar physical conditions as those present in the adjoining tidal flats, while the biology  
250 should also mirror that of a typical tidal flat habitat. This implies minimal mud content in the  
251 sediments, no mangrove root biomass, dissipation of inherited nutrients and no evidence of  
252 overabundant surface macroalgae. Importantly, macrofauna community structure should also be  
253 analogous to the adjacent tidal flat communities.

254 There are two main issues relating to monitoring of restoration/rehabilitation following mangrove  
255 clearing: firstly, the range of parameters that are monitored; and secondly the monitoring time  
256 frames. Monitoring of mangrove removal in New Zealand to date, has been variable in terms of  
257 temporal sampling, and as yet, there are no cases where repeated monitoring has continued beyond  
258 three years (e.g., Alfaro, 2010; Bulmer and Lundquist, 2014, 2015, 2016). Typically wetland  
259 restoration projects report some satisfactory level of recovery long after a 5 year timeframe,  
260 although this is usually in the context of planting for rehabilitation rather than clearance of  
261 vegetation. Siple and Donahue (2013) suggest that cleared sites in Hawaii may take decades to  
262 return to a pre(mangrove)-clearing/pre-invasion condition. Given the lack of available data in New  
263 Zealand, clearly any attempt at a synthesis of findings is going to be greatly limited. Stokes (2010)  
264 and Bulmer and Lundquist (2016) have reported minimal coarsening of sediment after 18 months to  
265 3 years, while other sites with more regular and stronger flow velocities achieved a more substantial  
266 coarsening (Stokes, 2009; Alfaro, 2010). Conversely, where cleared areas are large in relation to  
267 flushing potential, mud content has been observed to increase, at least within that 2 year timeframe  
268 (Bulmer and Lundquist, 2015).

269 Sediment condition drives benthic community structure (Cummings et al., 2003, Thrush et al., 2003).  
270 There is little evidence available to suggest that benthic communities within cleared substrates  
271 become consistent with tidal flat community structure within the first few years (Stokes, 2010;  
272 Lundquist et al., 2012; Bulmer and Lundquist, 2015). Outside of New Zealand, assessments of  
273 benthic food webs following clearance of red mangrove (*Rhizophora*) canopy, suggests that a  
274 recovery from removal occurs gradually (and is not governed by top-down effects (Siple and  
275 Donahue, 2013). An initial increase in species abundance, sometimes coupled with increased species  
276 diversity, is often noted, but assumed to be a response of opportunistic species to the liberation of  
277 benthic food sources (Felsing 2006, Alfaro 2010, Sweetman et al., 2010, Lundquist et al., 2012, Siple  
278 and Donahue 2013). Long term monitoring is required to separate seasonal fluctuations, the influx  
279 of short-term colonising species, and the development of stable benthic diversity. Again, this  
280 reinforces the necessity for a minimum of 5 years monitoring to confirm whether a return to tidal  
281 flat biogeochemistry and biological community structure occurs.

282 The monitoring parameters outlined in these case studies seem to have overlooked some potential  
283 unfavourable impacts of mangrove clearance. Typically these relate to the measure of  
284 biogeochemical change that develops on and below the surface. Firstly, surface topography is

285 typically not measured. Mangrove-dominated substrates consists of sediment fines mixed with  
286 large volumes of mangrove root material (Kauffman and Donato 2012). How quickly the surface  
287 topography changes in response to mangrove clearing will be partly linked to the existing  
288 belowground biomass (mangrove root material). The depth and density of mangrove root material,  
289 and its decomposition over time, is already typically ignored in monitoring schemes. Root material  
290 varies temporally and spatially (Saintilan, 1997; Komiyama et al., 2008; Lovelock, 2008), making it  
291 challenging to measure decomposition rates. Stokes and Harris (2015) described a considerable  
292 coarsening of sediment between 3 and 4 years post-clearance which they hypothesised may be  
293 linked to the slow rate of root decomposition. The timeframe sits within model predictions  
294 developed by (Gladstone-Gallagher et al., 2014) where buried wood and pneumatophores  
295 decompose to half their original weight between 317 and 613 days. Mangrove roots resist decay  
296 because they grow in saturated, low oxygen soils (Middleton and McKee, 2001) and their rate of  
297 decay is influenced by species (Middleton and McKee 2001), location and tidal elevation (Huxham et  
298 al., 2010). Generally, however, complete decay of woody material is expected to take several years  
299 (Mackey and Smail, 1996; Middleton and McKee, 2001; Siple and Donahue, 2013), potentially  
300 decades (Gladstone-Gallagher et al., 2014; Lundquist et al., 2014).

301 Another aspect of change in surface topography that is rarely included in monitoring of mangrove  
302 removals is that of site elevation. If sediments and biological material are flushed from a site, then a  
303 considerable deflation of the intertidal surface can be expected. This phenomenon has been  
304 reported in New Zealand, where surface elevation fell by 9 – 38 mm within 2 years of mangrove  
305 removal (Stokes et al., 2009). This is consistent with findings reported in other countries, where  
306 above-ground mangrove vegetation was cleared for development (Hayden and Granek, 2015), and  
307 where mangroves were destroyed during a hurricane (Cahoon et al., 2003). A lowering of surface  
308 levels has potential implications for nutrient loading and sedimentation on adjacent nearshore  
309 habitats. Such geomorphological change also results in an increase in relative sea level which may  
310 impact on adjacent saltmarsh habitat (Rogers et al., 2013; Saintilan and Rogers, 2013). Additionally,  
311 exposed shorelines may be more vulnerable to erosion, particularly if localised relative sea level rise  
312 is coupled with predicted global sea level rise (Hayden and Granek, 2015).

313 Finally, while a few consent monitoring schemes have looked at adverse impacts such as macro-algal  
314 blooms and bacterial blooms, in most cases in New Zealand, these have not been incorporated into  
315 monitoring designs. Granek and Ruttenberg (2008) suggest even small-scale clearing of mangroves  
316 can lead to changes in biotic conditions. Their study in the Caribbean identified a link between  
317 cleared sites and increased macro-algal biomass and cyanobacterial growth, both earlier than 12  
318 months and after 8 years. Strong links between nutrients and algal blooms exist in many estuaries  
319 (Anderson et al., 2002), particularly urbanised locations (Wallace and Gobler, 2014). Macroalgae  
320 blooms, specifically opportunistic Ulvacean species are a common symptom of eutrophication  
321 (Valiela et al., 1997). Macroalgal blooms may be driven by changes to the N:P or N:Si ratio, while  
322 shifts in macroalgae species are linked to an increase in the ratio of DOC:DON (Anderson et al.,  
323 2002). Measures of bacterial or algal biomass have not been part of monitoring protocols in New  
324 Zealand post mangrove removal, however significant macro-algal blooms associated with persistent  
325 mangrove debris were reported in Tauranga Harbour and elsewhere (Lundquist et al., 2014). It is  
326 important to have some knowledge of the background nutrient loading of an estuary in order to  
327 determine the likelihood of blooms following mangrove eradication.

328 It is important to highlight here two additional environmental benefits provided by mangroves that  
329 have not been considered in the development of mangrove management strategies in New Zealand.  
330 Firstly, mangroves are known to contribute to the health of coral reef and fisheries (Mumby and

331 Steneck, 2008), although these services may be less important in temperate than in tropical  
 332 mangrove forests (Morrisey et al., 2010). Reef filter feeders, for example, take up mangrove derived  
 333 nutrients; while mangrove organic matter can be transported further afield by fish movement (Lee,  
 334 1995). Mangroves may therefore be an important source of nutrients to adjacent reef systems and  
 335 therefore any reduction in mangroves may lead to reduced nutrient availability for sessile reef  
 336 organisms (Granek et al., 2009). A more recently recognised ecosystem benefit is that of climate  
 337 change mitigation. Numerous studies have identified the enormous carbon burial capacity of  
 338 mangroves (Kristensen et al., 2008). Any activity that impacts on carbon stores, such as the removal  
 339 of mangrove vegetation, and in turn adds to atmospheric carbon dioxide, can be viewed as a  
 340 negative outcome for climate change mitigation (Bulmer et al., 2015 and 2016). This aspect of  
 341 ecosystem management could also be considered in the context of mangrove removal in New  
 342 Zealand.

343

344 **4. Monitoring objectives to identify restoration goals following mangrove eradication**

345 Even though regulatory authorities today are often constrained by limited budgets and staff, an  
 346 opportunity exists to link industry needs with research objectives. Below, we provide a discussion  
 347 on what we consider to be key monitoring parameters. Collectively, these parameters can be used  
 348 to asses if and when management objectives are achieved. We have detailed the recommended  
 349 monitoring approaches using three levels (A, B and C in Table 2), grouped according to necessity,  
 350 complexity and expense (Table 2). Level A monitoring parameters reflect methods that can be  
 351 utilised with limited budgets, and can also be performed by community or volunteer groups with  
 352 some supervision of techniques and data reporting. Level B monitoring parameters are those that  
 353 would require some laboratory analysis and/or specialist knowledge (i.e. benthic macrofauna  
 354 analysis) while Level C monitoring parameters could be included in well-funded, comprehensive  
 355 monitoring programs. Information needs are compiled under three key themes; physical,  
 356 chemical and biological, detailed in Sections 4.1 – 4.3. Additionally, as levels of replication must be  
 357 considered when planning any monitoring program, we have included recommended levels of  
 358 replication relative to the size of proposed clearance areas (Figure 2).

359

360

361 Table 2. Hierarchical measurements suggested for monitoring of mangrove removals, as per details  
 362 in text. Details of the methodologies and limitations are described in Sections 4.1 – 4.3.

<b>MEASURING PHYSICAL &amp; CHEMICAL CHANGES (sampling intact mangroves; cleared; tidal flat)</b>		
<b>LEVEL A monitoring</b>	<b>LEVEL B monitoring</b>	<b>LEVEL C monitoring</b>
Description of surface sediment using field-based methods <a href="http://soilquality.org.au">http://soilquality.org.au</a> Sediment compaction (Schwarz et al., 2004 or Bulmer & Lundquist, 2014) Redox Potential Depth (Bulmer & Lundquist, 2014) Surface elevation – basic method (Schwarz et al., 2004) Bank erosion – photopoints (Schwarz et al., 2004)	Grain size: clay; silt, sand, gravel fractions Root biomass within macrofauna cores (Bulmer & Lundquist, 2014) Sediment organic content (Stokes, 2009) Grain size changes with depth using field-based methods (Stokes, 2009) Sediment chlorophyll-a content Sediment organic content (LOI method)	Surface elevation changes – long term RSET method (Stokes et al., 2009) Nutrient concentrations (N, P) Sediment accumulation in adjacent vegetated habitats (sediment traps; Stokes et al., 2010) Grain size changes with depth using lab-based analysis of size fractions Sediment dating – to develop mangrove sediment accretion history and rates of carbon accumulation Stable isotope analysis – to identify carbon sources

<b>MEASURING BIOLOGICAL CHANGES</b>		
LEVEL A monitoring	LEVEL B monitoring	LEVEL C monitoring
Habitat change: seagrass (Bulmer & Lundquist, 2014) Habitat change: saltmarsh (Bulmer & Lundquist, 2014) Habitat change: shellfish (Bulmer & Lundquist, 2014)	Habitat change: sampling established/pre-existing shellfish abundance	Habitat change: Shellfish condition – adjacent sandflat and established/pre-existing shellfish beds (Bulmer & Lundquist, 2014)
Surface observations <ul style="list-style-type: none"> <li>- Epifaunal species/abundance</li> <li>- No. of dead pneumatophores</li> <li>- Any new seeds/seedlings</li> <li>- % cover of macroalgae/algal blooms</li> <li>- Crab burrow abundance</li> <li>- Shallow infaunal bivalves (handraking upper 5 cm)</li> <li>- Depth to root mat (&amp; note presence/absence of root mass)</li> </ul>	Benthic macrofauna counts (0.5 mm sieve; Bulmer & Lundquist, 2014)  Bird monitoring (Schwarz et al., 2004)	

363

364

365

#### 366 **4.1 Monitoring physical parameters**

367

368 The following parameters must be considered (before and after mangrove removal) in order to  
369 develop a detailed understanding of the physical changes that occur within an estuary following  
370 mangrove removal;

- 371 a) Grain size change on the surface
- 372 b) Grain size change with depth (compaction would be associated with this)
- 373 c) Vertical elevation changes and change to relative sea level (tidal inundation)
- 374 d) Potential for bank erosion

375 Sediment grain size is a key measure of ‘recovery’, as benthic community structure is most likely to  
376 respond to changes in mud content (Thrush, 2003). Surface sediments should be sampled in order  
377 to identify which sediment size fraction is flushed following mangrove removal. The sampled depth  
378 can vary, although ideally this should be limited to the upper 2 – 5 mm. Results can be reported  
379 using % mud and % sand, via sieving methods, although a more useful characterisation of surface  
380 sediment might include % silt, % clay, and at least 3 sand fractions.

381 As sediments become more compacted, amenity value (i.e. walking access) increases. Sediment  
382 compaction is most easily monitored by assessing the depth to which footprints penetrate the  
383 surface (Bulmer and Lundquist, 2014). Commercial penetrometers can also be used, however for  
384 the sake of cost and simplicity, community groups generally utilise a steel rod with a pointed end.  
385 This is dropped from a known and consistent height and depth of penetration into the substrate is  
386 measured (Alfaro, 2010; Schwarz et al., 2004). Logic demands that as any remaining mangrove roots  
387 die and decompose, the surrounding sediments become softer and more penetrable, and  
388 conversely, as muds are flushed out of the system, sediments will compact and become firmer

389 underfoot. However, we recommend that sampling is augmented by a basic stratigraphical  
390 assessment, as sand armouring can camouflage persistent muds.

391 A recent study has identified the potential for sand armouring where mangroves are cleared in  
392 locations that receive low flow velocities (Stokes and Harris, 2015). This suggests that a reported  
393 positive change in surface grain size from mud dominated to sand dominated, may mask the  
394 trapping and retention of buried mud-dominated sediments. Simple stratigraphical descriptions  
395 could address this. A 30 cm core could be described using field-based textural assessments at 2 – 5  
396 cm intervals (see <http://soilquality.org.au/factsheets/soil-texture> as an example).

397 An important, but often overlooked impact of change following mangrove removal is the potential  
398 for significant lowering of the surface topography as a result of sediment flushing, sediment  
399 compaction and root collapse (Cahoon et al., 2003; Stokes et al., 2009). A lower seabed increases  
400 relative sea level, which, as a result of increased inundation may impose physiological stresses on  
401 adjoining saltmarsh communities or increase the risk of bank erosion. Schwarz et al., (2004) describe  
402 a low cost approach for monitoring bed-level whereby 1 m long wooden stakes are hammered into  
403 the sediment, along a transect. This technique was utilised by community groups to capture ‘surface  
404 elevation change’. One limitation of this approach is the unknown vertical movement of the stake  
405 which may artificially amplify results. Use of longer, thinner stainless steel pegs is desirable as these  
406 are more stable than wooden pegs, and being of smaller diameter, it is less likely that erosion will  
407 occur around the peg itself. Replication at each sampling station would be required, preferably with  
408 6 – 12 pegs positioned within a ½ m plot (Stokes, 2009). A more reliable (but equally more  
409 expensive) method employs Rod Sediment Elevation Tables (RSET) see (Stokes et al., 2009). These  
410 RSETs are driven many metres into the sediment and provide the opportunity for long term  
411 monitoring of inflation or deflation of the intertidal substrate (Cahoon et al., 2000).

#### 412 **4.2 Monitoring chemical parameters**

413 To develop an understanding of chemical changes in the substrate following mangrove removal we  
414 must consider:

- 415 a) Nutrient loading and algal bloom occurrence
- 416 b) Background nutrient loads
- 417 c) Oxic layer/redox

418 Estuaries have enormous capacity to accumulate sediments, nutrients and contaminants (Anderson  
419 et al., 2002). If a site already harbours high nitrogen and phosphorous levels, introducing  
420 decomposing mangrove litter, and liberating carbon rich mangrove sediments is likely to shift the  
421 system into a eutrophic state. Algal blooms are associated with nutrient rich systems, and can pose a  
422 real threat to estuarine organisms (Howarth et al., 2011). As such, baseline surveys need to include  
423 a spatial assessment of sediment nutrient loads, along with a record of the presence or absence of  
424 existing algal cover. Sampling of porewater nutrients can quantify whether removal areas have  
425 elevated nutrients, and whether these nutrients are likely transported into neighbouring habitats.  
426 Monitoring of temporal/seasonal variability in macro-algal blooms can determine how long-lasting  
427 nutrient impacts are following mangrove removal. In addition, nutrient sources can be determined,  
428 using elemental ratios (i.e. Yamamuro, 2000), which would identify sites loaded by mangrove detrital  
429 decomposition and/or sites smothered by macroalgae.

430 Measurement of sediment redox characteristics should also be included in any prescribed  
431 monitoring, as the development of oxic sediments is a requirement for recolonization of sand-  
432 dwelling benthos. Measurements can be made via direct redox probe readings, or simply by

433 measuring the depth to which black/dark grey sediments occur (reflecting the end of the oxic layer),  
434 as described in Lundquist et al. (2012).

### 435 **4.3 Monitoring biological parameters**

436 To develop an understanding of biological changes following mangrove removal we must consider:

- 437 a) Benthic fauna
- 438 b) Vegetative condition of clearance sites
- 439 c) Impact on neighbouring habitats (e.g. bivalve beds, seagrass meadows, salt marsh)

440 Macroinvertebrates are often used as an indicator of environmental change (Hilty and Merenlender,  
441 2000). As surface fauna (i.e. epifauna), are easily identified and counted, this approach has been  
442 incorporated into both community and consented monitoring. Limited species knowledge of  
443 epifaunal taxa is required as only 4 to 6 grazing species are typically encountered in visual surveys.  
444 In addition, hand raking for bivalves is a rapid sampling method, and generates important  
445 information on the recovery status of the site. Conversely, enumerating other buried  
446 macroinvertebrates, or infauna, is a time-intensive process that often requires specific taxonomic  
447 expertise, particularly to identify the numerous polychaete species typically associated with  
448 mangrove and tidal flat habitats. Assessments of species abundance and richness of benthic  
449 macroinvertebrates is commonly used to assess change; however caution must be observed when  
450 interpreting results as these univariate metrics document only number of species or individuals,  
451 which may not reflect successful restoration of ecological function. Rather high values of these  
452 univariate metrics may instead reflect macrofaunal communities dominated by opportunistic or  
453 disturbance tolerant species, which has often been documented in degraded systems typical of  
454 unsuccessful mangrove removals (Lundquist et al., 2012, 2014). Multivariable statistical approaches  
455 are a preferred method to analyse restoration success of benthic macrofaunal communities, and  
456 allow differentiation between communities dominated by ephemeral, opportunistic colonisers from  
457 species that are indicative of a rehabilitated site.

458 As fish and shorebirds (at least in temperate mangroves in New Zealand) tend to only spend small  
459 proportions of the tidal cycle within mangrove ecosystems, typically they are poor indicators of  
460 restoration success unless substantial efforts are put into elucidating spatial and temporal (intra-  
461 and inter-annual) patterns in fish and bird usage of mangrove habitats. To date, information to  
462 confirm the relative role of mangroves for key species is not yet available, though monitoring in  
463 Whangamata Harbour of bird responses to mangrove removals suggests minimal impacts on birds  
464 after removals, where mangrove habitats remain (Wildlands, 2014).

465 The presence of mangrove root biomass is another key measure for determining whether a site is  
466 transitioning towards tidal flat conditions, as thick mangrove biomass delays both sediment erosion  
467 and colonisation by macrofauna. This can be rapidly assessed by counting the number of  
468 pneumatophores (if present), and digging into the sediment to detect the presence of a root mass  
469 and quantify volumes.

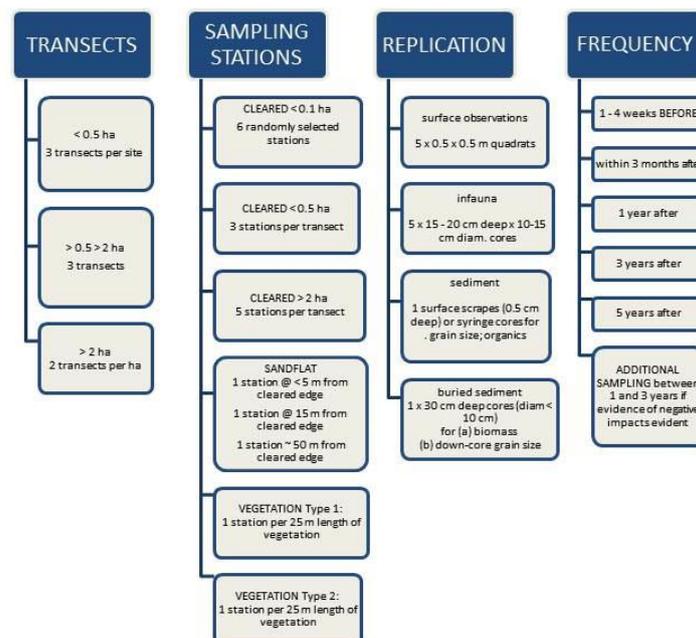
470 Monitoring should include assessment of potential impacts on neighbouring habitats, including  
471 habitats of particular importance such as shellfish beds and seagrass meadows. If the size of a  
472 mangrove clearing is large compared to the expected flushing of sediment and nutrients, it is likely  
473 that adverse downstream effects on sand-dwelling macroinvertebrates will occur. However, note  
474 that to date, no adverse impacts have been detected on neighbouring tidal flat and seagrass  
475 meadows, even within locations within a few metres of mangrove removals. This is likely due to the  
476 slow rates of sediment erosion observed to date from mangrove removals, such that rates of

477 sediment erosion have been unlikely to impact on neighbouring habitats. Bivalves in particular are  
 478 an important component of the estuarine trophic system, providing food for foraging birds, fish and  
 479 people. Some bivalves, such as the New Zealand species *Paphies australis* and *Macomona liliiana*  
 480 prefer a sand-dominated substrate and show adverse sensitivities to an increase in mud content.  
 481 Similarly, redistribution of fine sediments may lead to increased turbidity and sediment deposition,  
 482 and decline of seagrass, while landward transport of sediment may smother remaining mangroves or  
 483 saltmarsh beyond their ability to thrive. Baseline surveys are a necessity, followed by seasonal  
 484 observations of plant species richness, density and general condition.

485 In New Zealand, early attempts at mangrove clearing were restricted to clearing above-ground  
 486 biomass by hand from narrow marginal strips. More recently however, large machinery in the form  
 487 of wide-track diggers, have been used to cut and mulch larger areas of mangrove vegetation  
 488 (Lundquist et al., 2014). Observed dieback of mangroves adjacent to some of the cleared plots  
 489 suggests that large-scale mechanical clearing may be detrimental to neighbouring vegetation. Visual  
 490 descriptions, coupled with repeated photographic documentation from nominated 'photo-points',  
 491 are a useful method of picking up any changes to the coverage and condition of remaining  
 492 vegetation. Along with mangroves, this method can also include an assessment of seagrass and  
 493 saltmarsh; these techniques have also been used to measure rates of encroachment of mangrove  
 494 forests into saltmarsh habitats, a process that is anticipated to increase in frequency with sea level  
 495 rise.

#### 496 4.4 Replication and frequency of sampling

497 It is imperative that any restoration program include regular monitoring at an appropriate scale of  
 498 replication to document rates of recovery and any adverse or unexpected impacts. Lundquist and  
 499 Bulmer (2015) documented little change in some monitoring parameters within 12 months of  
 500 mangrove removal in Whangamata. As such, a more staggered repetition of monitoring may be  
 501 required to ensure longer-term sampling can be incorporated. Figure 2 provides a suggested  
 502 sampling protocol relative to the size of the area to be cleared of mangroves.  
 503



505 **Figure 2. Suggested levels of sampling replication, relative to size of area subjected to mangrove**  
506 **clearing.**

507

## 508 **5. Final remarks**

509 The suggested monitoring parameters discussed above not only hint at the complexity of estuarine  
510 systems but also highlight the need for detailed before, after and repeated monitoring programs  
511 where mangrove removal occurs. Unfortunately the mangrove removal programs in New Zealand  
512 have not yet led to the development of a consistent management or monitoring approach. Indeed,  
513 there is little conclusive evidence that any key management objective has been achieved. Resource  
514 managers must first have a clear management objective in mind (e.g. whether a site is to be  
515 “restored” to a pre-determined pristine condition or to one that is “fit for purpose” (not necessarily  
516 pristine), and in so doing seek a better understanding of a) whether stressors associated with  
517 mangrove removal can be stopped or mitigated and b) whether the system will recover on its own or  
518 whether continued intervention is required (Elliot et al., 2007). The background, pre-removal  
519 conditions of an estuary will influence the rate of restoration. Detailed, repeated monitoring is  
520 central to the development of an informed management program designed to support a healthy  
521 estuarine ecosystem.

### ***Acknowledgements***

The first author would like to thank NIWA Hamilton for use of office space to complete this manuscript. Also, we are grateful to Dr Catherine Beard for clear and useful feedback on earlier drafts.

## References

- Alfaro, A. C., 2010. Effects of mangrove removal on benthic communities and sediment characteristics at Mangawhai Harbour, northern New Zealand. *Ices Journal of Marine Science*, 67, 1087-1104.
- Anderson, D. M., P. M. Glibert & Burkholder, J. M., 2002 Harmful algal blooms and eutrophication: Nutrient sources, composition, and consequences. *Estuaries*, 25, 704-726.
- Barbier, E. B., S. D. Hacker, C. Kennedy, E. W. Koch, A. C. Stier & Silliman B. R., 2011. The value of estuarine and coastal ecosystem services. *Ecological Monographs*, 81, 169-193.
- Bayraktarov, E., Saunders, M.I., Abdullah, S., Mills, M., Beher, J., Possingham, H.P., Mumby, P.J. and Lovelock C.E., 2015. The cost and feasibility of marine coastal restoration. *Ecological Applications*.
- Beard, C., 2006. Physiological constraints on the latitudinal distribution of the mangrove *Avicennia marina* (Forsk.) Vierh. subsp. *australasica* (Walp.) J. Everett in New Zealand. Hamilton, New Zealand: University of Waikato, Doctoral thesis, 203p.
- Bulmer, R. & Lundquist, C., 2014. Whangamata mangrove removal monitoring. Summary of 12 month post-removal sampling. NIWA Technical Report. Prepared for Waikato Regional Council. NIWA, Hamilton, New Zealand.
- Bulmer, R. & Lundquist C.J., 2015. Whangamata mangrove removal monitoring. Summary of 24 month post-removal sampling. NIWA Technical Report. Prepared for Waikato Regional Council. NIWA, Hamilton, New Zealand.
- Bulmer, R. & Lundquist, C.J., 2016. Whangamata mangrove removal monitoring. Summary of 36 month post-removal sampling. NIWA Technical Report. Prepared for Waikato Regional Council. NIWA, Hamilton, New Zealand.
- Bulmer, R.H., Lundquist, C.J., Schwendenmann, L., 2015. Sediment properties and CO<sub>2</sub> efflux from intact and cleared temperate mangrove forests. *Biogeosciences*, 12(20): 6169-6180.
- Bulmer, R.H., Schwendenmann, L., Lundquist, C.J., 2016. Carbon and nitrogen stocks and below-ground allometry in temperate mangroves. *Frontiers in Marine Science*, 3: 150.
- Cahoon, D. R., P. Hensel, J. Rybczyk, K. L. McKee, C. E. Proffitt & Perez, B. C., 2003. Mass tree mortality leads to mangrove peat collapse at Bay Islands, Honduras after Hurricane Mitch. *Journal of Ecology*, 91, 1093-1105.
- Cahoon, D. R., P. E. Marin, B. K. Black & Lurch, J. C., 2000. A method for measuring vertical accretion, elevation, and compaction of soft, shallow-water sediments. *Journal of Sedimentary Research*, 70, 1250-1253.
- Chapman, V.J., 1976a. Mangroves and salt marshes of the Kaipara Harbour: a study with proposals for preservation of areas supporting the harbour ecosystem. Department of Lands and Survey, Auckland, 28 p.
- Chapman, V.J., 1976b. Mangroves and salt marshes of the Herekino, Whangape and Hokianga Harbours: a study with proposals for preservation of areas supporting the harbour ecosystem. Department of Lands and Survey, Auckland, 28 p.
- Cummings, V., S. Thrush, J. Hewitt, A. Norkko & Pickmere, S., 2003. Terrestrial deposits on intertidal sandflats: sediment characteristics as indicators of habitat suitability for recolonising macrofauna. *Marine Ecology Progress Series*, 253, 39-54.

- Dale, P. E. R., Knight, J. M., & Dwyer, P.G., 2014. Mangrove rehabilitation: a review focusing on ecological and institutional issues. *Wetlands Ecology and Management*, 22(6), 587-604.
- de Lange, W. & de Lange, P.J., 1994. An appraisal of factors controlling the latitudinal distribution of Mangrove (*Avicennia marina* var. *resinifera*) in New Zealand. *Journal of Coastal Research*, 10, 539-548.
- de Luca, S., 2015. Mangroves in NZ - misunderstandings and management [online]. In: Australasian Coasts & Ports Conference 2015: 22nd Australasian Coastal and Ocean Engineering Conference and the 15th Australasian Port and Harbour Conference. Auckland, New Zealand: Engineers Australia and IPENZ, 2015: 242-245.
- Doughty, C., J. A. Langley, W. Walker, I. Feller, R. Schaub & Chapman, S., 2015. Mangrove Range Expansion Rapidly Increases Coastal Wetland Carbon Storage. *Estuaries and Coasts*, 1-12.
- Elliott, M., D. Burdon, K. L. Hemingway & Apitz, S. E., 2007. Estuarine, coastal and marine ecosystem restoration: Confusing management and science - A revision of concepts. *Estuarine Coastal and Shelf Science*, 74, 349-366.
- Ellis, J., Nicholls, P., Craggs, R., Hofstra, D., & Hewitt J., 2004. Effects of terrigenous sedimentation on mangrove physiology and associated macrobenthic communities. *Marine Ecology Progress Series* 270, 71-82.
- Ellison, A. M., 2008. Managing mangroves with benthic biodiversity in mind: Moving beyond roving banditry. *Journal of Sea Research*, 59, 2-15.
- Felsing, M., 2006. Memo: Benthic impacts of mangrove removal Whangamata harbour - analysis of Nov 05 samples. Environment Waikato.
- Ferrier, R. C. & Jenkins A., 2010. Handbook of catchment management. Wiley-Blackwell.
- Giri, C., E. Ochieng, L. L. Tieszen, Z. Zhu, A. Singh, T. Loveland, J. Masek & Duke, N., 2011. Status and distribution of mangrove forests of the world using earth observation satellite data. *Global Ecology and Biogeography*, 20, 154-159.
- Gladstone-Gallagher, R. V., C. J. Lundquist & Pilditch, C.A., 2014. Mangrove (*Avicennia marina* subsp *australasica*) litter production and decomposition in a temperate estuary. *New Zealand Journal of Marine and Freshwater Research*, 48, 24-37.
- Granek, E. & Ruttenberg, B. I., 2008. Changes in biotic and abiotic processes following mangrove clearing. *Estuarine Coastal and Shelf Science*, 80, 555-562.
- Granek, E. F., Compton, J. E., & Phillips, D.L., 2009. Mangrove-exported nutrient incorporation by sessile coral reef invertebrates. *Ecosystems*, 12(3), 462-472.
- Halpern, B. S., K. L. McLeod, A. A. Rosenberg & Crowder, L. B., 2008. Managing for cumulative impacts in ecosystem-based management through ocean zoning. *Ocean & Coastal Management*, 51, 203-211.
- Harty, C., 2009. Mangrove planning and management in New Zealand and South East Australia—A reflection on approaches. *Ocean & Coastal Management*, 52 (5), 278-286.
- Harrison Grierson Consultants Limited, 2012. Resource Consent Application, removal and management of mangroves – Tauranga Harbour. Document Number R001v1-TG133102-01.
- Hayden, H. L. & Granek, E. F., 2015. Coastal sediment elevation change following anthropogenic mangrove clearing. *Estuarine Coastal and Shelf Science*, 165, 70-74.

- Hilty, J. & Merenlender, A., 2000. Faunal indicator taxa selection for monitoring ecosystem health. *Biological conservation*, 92, 185-197.
- Howarth, R., F. Chan, D. J. Conley, J. Garnier, S. C. Doney, R. Marino & Billen, G., 2011. Coupled biogeochemical cycles: eutrophication and hypoxia in temperate estuaries and coastal marine ecosystems. *Frontiers in Ecology and the Environment*, 9, 18-26.
- Huxham, M., J. Langat, F. Tamoooh, H. Kennedy, M. Mencuccini, M. W. Skov & Kairo, J., 2010. Decomposition of mangrove roots: Effects of location, nutrients, species identity and mix in a Kenyan forest. *Estuarine, Coastal and Shelf Science*, 88, 135.
- Kauffman, J. B. & Donato, D. C., 2012. Protocols for the measurement, monitoring and reporting of structure, biomass and carbon stocks in mangrove forests. In Working Paper 86. Bogor, Indonesia: CIFOR.
- Kennish, M. J., 2002. Environmental threats and environmental future of estuaries. *Environmental Conservation*, 29, 78-107.
- Komiyama, A., J. E. Ong & Pongpan, S., 2008. Allometry, biomass, and productivity of mangrove forests: A review. *Aquatic Botany*, 89, 128.
- Kristensen, E., Bouillon, S., Dittmar, T., & Marchand, C., 2008. Organic carbon dynamics in mangrove ecosystems: a review. *Aquatic Botany*, 89(2), 201-219
- Lee, S.Y., 1995. Mangrove outwelling: a review. *Hydrobiologia*, 295:203–12.
- Lotze, H. K., H. S. Lenihan, B. J. Bourque, R. H. Bradbury, R. G. Cooke, M. C. Kay, S. M. Kidwell, M. X. Kirby, C. H. Peterson & Jackson, J. B. C., 2006. Depletion, degradation, and recovery potential of estuaries and coastal seas. *Science*, 312, 1806-1809.
- Lovelock, C., 2008. Soil respiration and belowground carbon allocation in mangrove forests. *Ecosystems*, 11, 342-354.
- Lundquist, C., S. Hailes, K. Cartner, K. Carter & Gibbs, M., 2012. Physical and ecological impacts associated with mangrove removals using in situ mechanical mulching in Tauranga Harbour. NIWA Technical Report No. 137. 106. Wellington, New Zealand.
- Lundquist, C. J., D. J. Morrissey, R. V. Gladstone-Gallagher & Swales, A., 2014. Managing mangrove habitat expansion in New Zealand. In *Mangrove Ecosystems of Asia*, ed. I. Faridah-Hanum, Latiff, A., Hakeem, K.R., Ozturk, M., 415-434. New York: Springer Science + Business Media.
- Mackey, A. P. & Smail, G., 1996. The decomposition of mangrove litter in a subtropical mangrove forest. *Hydrobiologia*, 332, 93-98.
- Middleton, B. A. & McKee, K. L., 2001. Degradation of mangrove tissues and implications for peat formation in Belizean island forests. *Journal of Ecology*, 89, 818-823.
- Milbrandt, E. C., M. Thompson, L. D. Coen, R. E. Grizzle & Ward, K., 2015. A multiple habitat restoration strategy in a semi-enclosed Florida embayment, combining hydrologic restoration, mangrove propagule plantings and oyster substrate additions. *Ecological Engineering*, 83, 394-404.
- Mildenhall, D.C., 2001. Middle Holocene mangroves in Hawkes Bay. *NZ Journal of Botany*, 39, 517-521.
- Morrissey, D. J., A. Swales, S. Dittmann, M. A. Morrison, C. E. Lovelock & Beard, C. M., 2010. The Ecology And Management Of Temperate Mangroves. In *Oceanography And Marine Biology: An Annual Review*, Vol 48, 43-160. Boca Raton: Crc Press-Taylor & Francis Group.

- Mumby, P. J., & Steneck, R.S., 2008. Coral reef management and conservation in light of rapidly evolving ecological paradigms. *Trends in ecology & evolution*, 23(10), 555-563.
- Osland, M. J., A. C. Spivak, J. A. Nestlerode, J. M. Lessmann, A. E. Almario, P. T. Heitmuller, M. J. Russell, K. W. Krauss, F. Alvarez, D. D. Dantin, J. E. Harvey, A. S. From, N. Cormier & Stagg, C. L., 2012. Ecosystem Development After Mangrove Wetland Creation: Plant–Soil Change Across a 20-Year Chronosequence. *Ecosystems*, 15, 848-866.
- Park, S. 2004. Aspects of Mangrove Distribution and Abundance in Tauranga Harbour. Environment BOP Environmental publication 2004/16. 49. Whakatane.
- Rogers, K., N. Saintilan & Copeland, C., 2013. Managed Retreat of Saline Coastal Wetlands: Challenges and Opportunities Identified from the Hunter River Estuary, Australia. *Estuaries and Coasts*, 1-12.
- Rogers, K., N. Saintilan & Heijnis, H., 2005. Mangrove Encroachment of Salt Marsh in Western Port Bay, Victoria: the Role of Sedimentation, Subsidence and Sea Level Rise. *Estuaries*, 28, 551-559.
- Saintilan, N., 1997. Above- and below-ground biomasses of two species of mangrove on the Hawkesbury River estuary, New South Wales. *Marine and Freshwater Research*, 48, 147-152.
- Saintilan, N. & Rogers, K., 2013. The significance and vulnerability of Australian saltmarshes: Implications for management in a changing climate. *Marine and Freshwater Research*, 64, 66-79.
- Saintilan, N., N. C. Wilson, K. Rogers, A. Rajkaran & Krauss, K. W., 2014. Mangrove expansion and salt marsh decline at mangrove poleward limits. *Global Change Biology*, 20, 147-157.
- Schwarz, A., B. Burns & Alfaro, A.C., 2004. Guidelines for community-focused ecological monitoring of mangrove habitats in estuaries. Prepared for Environment Waikato. 25. Hamilton (New Zealand).
- Siple, M. C. & Donahue, M.J., 2013. Invasive mangrove removal and recovery: Food web effects across a chronosequence. *Journal of Experimental Marine Biology and Ecology*, 448, 128-135.
- Stokes, D. J., 2009. Assessment of physical changes following mangrove removal: Whangamata Harbour 2008. E.W. Technical Report 2009(13). Hamilton, New Zealand: Environment Waikato.
- Stokes, D. J., 2010. The physical and ecological impacts of mangrove expansion and mangrove removal: Tauranga Harbour, New Zealand. In *Earth Sciences*, 191. Hamilton, New Zealand: University of Waikato.
- Stokes, D. J., & Harris, R.J., 2015. Sediment properties and surface erodibility following a large-scale mangrove (*Avicennia marina*) removal. *Continental Shelf Research*, 107, 1-10.
- Stokes, D. J., T. R. Healy & Cooke, P. J., 2009. Surface elevation changes and sediment characteristics of intertidal surfaces undergoing mangrove expansion and mangrove removal, Waikaraka Estuary, Tauranga Harbour, New Zealand. *International Journal of Ecology & Development*, 12.
- Stokes, D. J., Healy, T.R. & Cooke, P.J., 2010. Expansion Dynamics of Monospecific, Temperate Mangroves and Sedimentation in Two Embayments of a Barrier-Enclosed Lagoon, Tauranga Harbour, New Zealand. *Journal of Coastal Research*, 26, 113-122.
- Swales, A., S. J. Bentley & Lovelock, C. E., 2015. Mangrove-forest evolution in a sediment-rich estuarine system: opportunists or agents of geomorphic change? *Earth Surface Processes and Landforms*, 40, 1672-1687.
- Swales, A., Bentley, S.J., Lovelock, C., & Bell, R.G., 2007. Sediment processes and mangrove-habitat expansion on a rapidly-prograding muddy coast, New Zealand. *Coastal Sediments*, 7, 1441–1454.

- Sweetman, A. K., J. J. Middelburg, A. M. Berle, A. F. Bernardino, C. Schander, A. W. J. Demopoulos & Smith, C. R., 2010. Impacts of exotic mangrove forests and mangrove deforestation on carbon remineralization and ecosystem functioning in marine sediments. *Biogeosciences*, 7, 2129-2145.
- Thrush, S. F., J. E. Hewitt, A. Norkko, P. E. Nicholls, G. A. Funnell & Ellis, J. I., 2003. Habitat change in estuaries: predicting broad-scale responses of intertidal macrofauna to sediment mud content. *Marine Ecology-Progress Series*, 263, 101-112.
- Thrush, S. F., M. Townsend, J. E. Hewitt, K. Davies, A. M. Lohrer, C. Lundquist & Cartner, K., 2013. The many uses and values of estuarine ecosystems. *Ecosystem services in New Zealand—conditions and trends*. Manaaki Whenua Press, Lincoln, New Zealand.
- Valiela, I., J. McClelland, J. Hauxwell, P. J. Behr, D. Hersh & Foreman, K., 1997. Macroalgal blooms in shallow estuaries: Controls and ecophysiological and ecosystem consequences. *Limnology and Oceanography*, 42, 1105-1118.
- Wallace, R. B. & Gobler, C. J., 2015. Factors Controlling Blooms of Microalgae and Macroalgae (*Ulva rigida*) in a Eutrophic, Urban Estuary: Jamaica Bay, NY, USA. *Estuaries and Coasts*, 38, 519-533.
- Wildlands, 2014. Monitoring of banded rail and other avifauna before and after mangrove clearance at Whangamata Harbour – Annual report (March 2013-2014). Contract report No. 3120b, prepared for Waikato Regional Council.
- Yamamuro, M., 2000. Chemical tracers of sediment organic matter origins in two coastal lagoons. *Journal of Marine Systems*, 26, 127-134.
- Zedler, J. B., J. M. Doherty & Miller, N. A., 2012. Shifting Restoration Policy to Address Landscape Change, Novel Ecosystems, and Monitoring. *Ecology and Society*, 17.
- Zhao, Q., J. Bai, L. Huang, B. Gu, Q. Lu & Gao, Z., 2016. A review of methodologies and success indicators for coastal wetland restoration. *Ecological Indicators*, 60, 442-452.